

***Aspergillus* spp. และสารพิษจากเชื้อราที่เกี่ยวข้องกับอาหารโคนม: อะฟลาทอกซินและกลีโอทอกซิน**
(*Aspergillus* spp. and mycotoxins associated with dairy cattle feeds: aflatoxins and gliotoxin)



โดย

นายอนุสรณ์ อยู่เย็น

นักวิทยาศาสตร์การแพทย์ชำนาญการพิเศษ
กลุ่มพิษวิทยาและชีวเคมี
สถาบันสุขภาพสัตว์แห่งชาติ



Research team

- ❖ **Anusorn Yooyen**
- ❖ **Assoc. Prof. Dr.Amnart Paopolathep**
- ❖ **Dr.Phattarawadee Wattanasuntorn**
- ❖ **Nuttakorn Ratchabut**
- ❖ **Dr.Muncharee Tattiyapong**
- ❖ **Phurida Sripipattanakul**
- ❖ **Onin Dejyong**

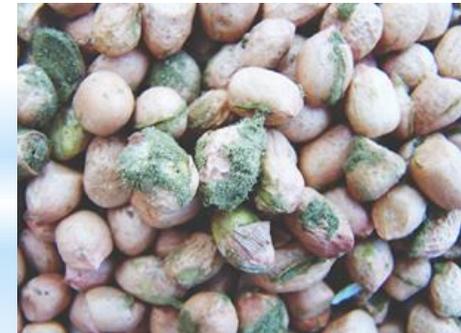
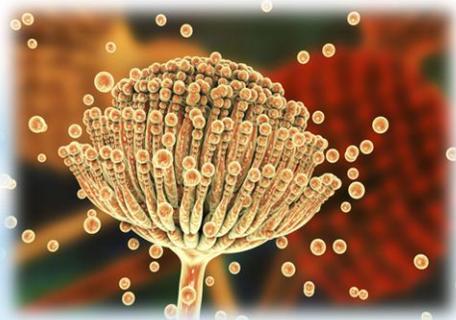
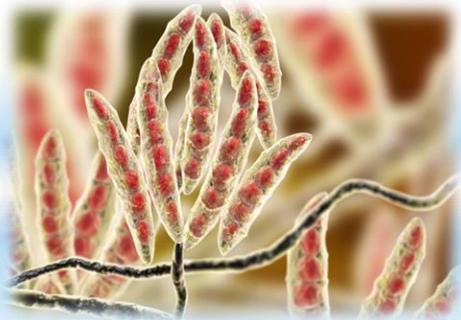


Introductions



Mycotoxins

- ❖ natural compounds produced as secondary metabolites
- ❖ filamentous fungi (*Aspergillus*, *Penicillium*, and *Fusarium*)
- ❖ contaminate around 25% of global food crops
- ❖ actual contamination rate can range from 60.0% to 80.0%



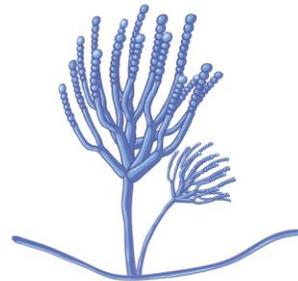
Mycotoxins (*cont.*)

- ❖ **Toxin-producing fungi can be divided into two categories:**
 - **Field fungi (e.g., *Fusarium* spp.)**
 - **Storage fungi (e.g., *Aspergillus* spp., *Penicillium* spp.)**

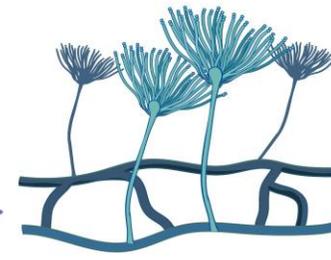
The different types of Mold



Bread mould



Penicillium



Aspergillus



Mycotoxins (*cont.*)

❖ Major mycotoxins

- **Aflatoxins (*Aspergillus flavus, A. parasiticus*)**
- **Fumonisin (*Fusarium verticillioides, F. proliferatum*)**
- **Ochratoxins (*Aspergillus ochraceus, A. carbonarius, Penicillium verrucosum*)**
- **Zearalenone (*Fusarium graminearum*)**
- **Trichothecenes (*Fusarium graminearum, F. sporotrichioides, F. poae*)**
- **emerging mycotoxins (beauvericin and enniatins) *F. acuminatum, F. avenaceum***



Aflatoxin (AFs)

❖ **Group of secondary metabolites produced mainly by fungal species such as**

- *Aspergillus flavus*

- *A. parasiticus*

- *A. nomius* (rarely)

❖ **Under humid conditions, these fungi are produced in**

- **maize, cottonseed, peanuts, tree nuts (almonds, pistachios, walnuts, etc.)**

- **livestock feed**

- **medicinal herbs**



Aflatoxin (*cont.*)

❖ Aflatoxin contamination

- ✓ can occur at any stage of food production

- ✓ from pre-harvest to storage stages of food chain

❖ Aflatoxin accumulation is dependent on

- Environmental such as:

- ✓ moisture

- ✓ temperature

- ✓ plant density

- Poor harvest practices

- Improper grain storage



Aflatoxin (*cont.*)

- ❖ **The removal of aflatoxins is very difficult due to their**
 - ✓ **stability and thermal resistance**
 - **melting point $>250^{\circ}\text{C}$**
 - **stable at a pH 3-10**
 - ✓ **resistant to food processing**
- ❖ **Thus remain unchanged throughout the food chain**



Classification of Aflatoxins

❖ There are more than 20 known aflatoxins, but the four main ones are

✓ Aflatoxin B₁ (AFB₁)

AFs B series are produced by

✓ *A. flavus*, *A. parasiticus* and *A. nomius*

✓ Aflatoxin B₂ (AFB₂)

✓ Aflatoxin G₁ (AFG₁)

AFs G series are produced by

✓ *A. parasiticus* and *A. nomius*

✓ Aflatoxin G₂ (AFG₂)

❖ Named based on their

✓ blue (B) or green (G) fluorescence under UV-light

✓ Relative mobility, 1 and 2 (higher and lower respectively) by TLC



Classification of Aflatoxins....

❖ Other members of the aflatoxin family

□ Aflatoxin M₁ (AFM₁) and Aflatoxin M₂ (AFM₂)

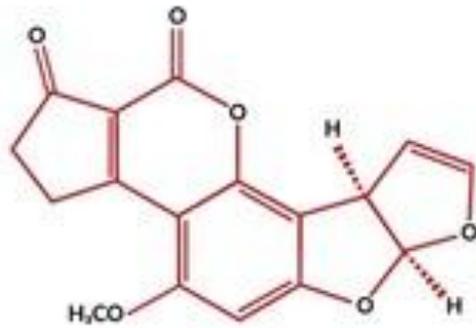
- ✓ are oxidative forms of Aflatoxin B₁ (AFB₁)
- ✓ modified in digestive tract of animals
- ✓ isolated from milk, urine and feces

❖ The level of toxicity associated aflatoxin varies the types present

- ✓ AFB₁ > AFG₁ > AFB₂ > AFG₂



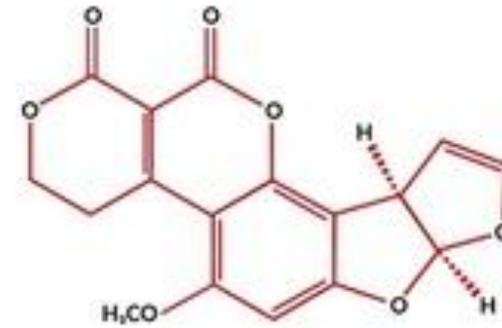
Chemical structure of aflatoxins



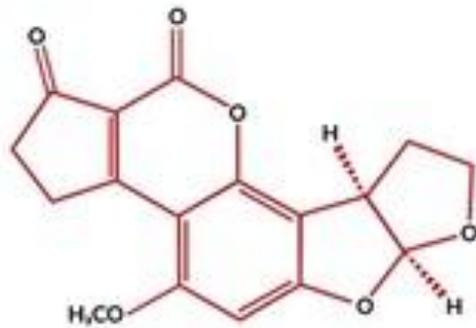
Aflatoxin B₁



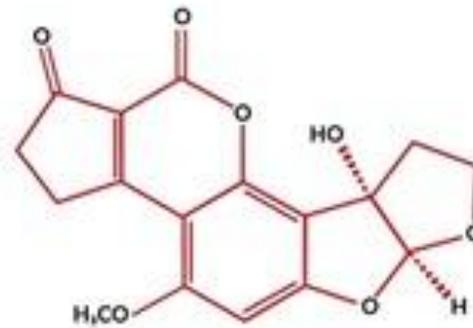
Aflatoxin M₁



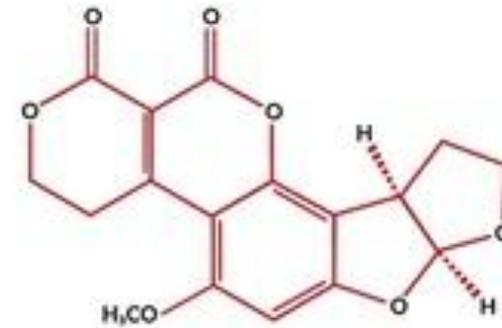
Aflatoxin G₁



Aflatoxin B₂



Aflatoxin M₂



Aflatoxin G₂



Aflatoxicosis in Animals

The toxic effect can include

- ❖ **Mutagenesis**
- ❖ **Carcinogenesis**
- ❖ **Teratogenesis**
- ❖ **Immunosuppression**
- ❖ **Hepatotoxicity (Liver damage with reduced protein synthesis)**
- ❖ **feed inappetence, poor growth rate, unthriftiness, depression, hemorrhage, icterus, and death**

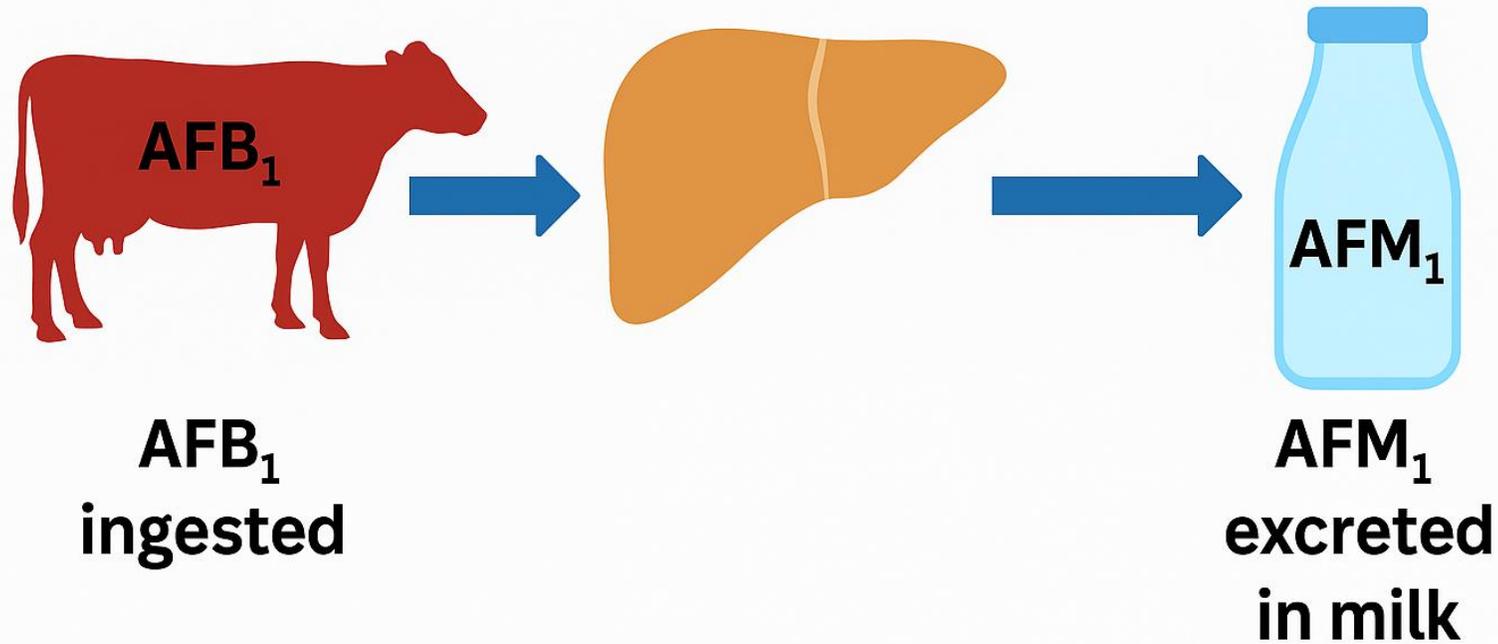


Health Effects in Dairy Cattle

- ❖ **Reduced feed intake**
- ❖ **Decreased milk production**
- ❖ **Immune suppression and increased disease susceptibility**
- ❖ **Liver damage and reproductive disorders.**
- ❖ **Oral LD₅₀ values for AFB₁ in young calves = 0.5-1 mg/kg**
- ❖ **Oral LD₅₀ values of AFB₁ estimates range from 0.3-17.9 mg/kg for many animals**



Biotransformed in liver



IARC has classified **AFB₁** and **AFM₁** as human carcinogens (IARC Group 1).



Gliotoxin (GT)

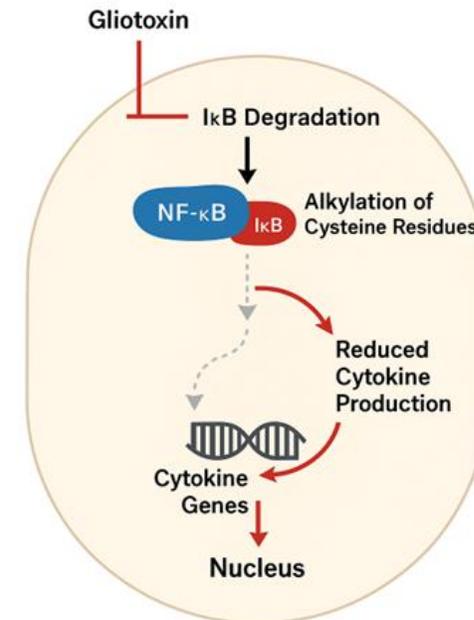
- ❖ secondary fungal metabolite
- ❖ Gliotoxin-Producing Fungi (*Aspergillus fumigatus*, *A. flavus*, *Eurotium chevalieri*, *Trichoderma virens*)
- ❖ Contaminate animal feed; complete feed (soybean meal, ground corn, and corn cobs, etc.)
- ❖ Exhibits immunosuppressive, cytotoxic, and genotoxic effects.



Immunosuppressive Effects

- ❖ **Inhibits NF- κ B activation** → **reduced cytokine production**
- ❖ **Impairs lymphocyte activation and defense mechanisms**

Immunosuppressive Effects of Gliotoxin



Cytotoxicity and Oxidative Stress

- ❖ Produces reactive oxygen species (ROS)
- ❖ Induces apoptosis via mitochondrial damage

Effects in Dairy Cattle

- ❖ Reduced feed intake and milk yield
- ❖ Increased risk of mastitis due to immunosuppression
- ❖ Liver damage; possible mycotoxin residues in milk
- ❖ General Signs: Depression, weakness, rough hair coat, poor growth in young calves, dehydration and diarrhea (if co-exposed with enterotoxins)



Oral LD₅₀ values for GT in mice = 67 mg/kg

Problem Statement and Rationale

- ❖ **Global Concern: Mycotoxin contamination poses serious risks to animal health and causes economic losses worldwide.**
- ❖ **Research Gap in Thailand: Most studies focus only on aflatoxins (AFs); few investigate co-contamination with gliotoxin (GT).**
- ❖ **Importance of Dual Detection: Animal feeds are often contaminated with multiple mycotoxins → increased health risks.**
- ❖ **Need for Improved Methods: LC/MS/MS requires validation for reliable detection of AFs and GT.**
- ❖ **Study Significance: Survey AFs and GT in dairy feeds, identify *Aspergillus* spp., and develop validated detection method.**



Objectives

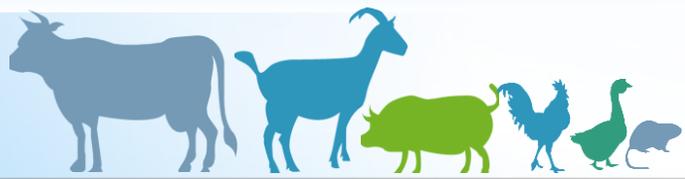
- ❖ To investigate the contamination of aflatoxins (AFB1, AFB2, AFG1, AFG2) and gliotoxin (GT) produced by fungi in the genus *Aspergillus* spp. in dairy cattle feed samples collected from farms in the Bangkok metropolitan area.
- ❖ To examine the molecular characteristics of fungi in the genus *Aspergillus* and investigate the genetic relationships among the isolates in correlation with the levels of aflatoxins (AFs) and gliotoxin (GT) detected.
- ❖ To develop and validate a method for the detection and quantification of aflatoxins (AFs) and gliotoxin (GT) in dairy feed samples using LC/MS/MS technique.



Materials and Methods



Part I: Fungal Isolation & Molecular characteristics of fungi



Sample collection



10 complete feeds



10 corn husks



20 Others

Fungal isolation

10 complete feeds



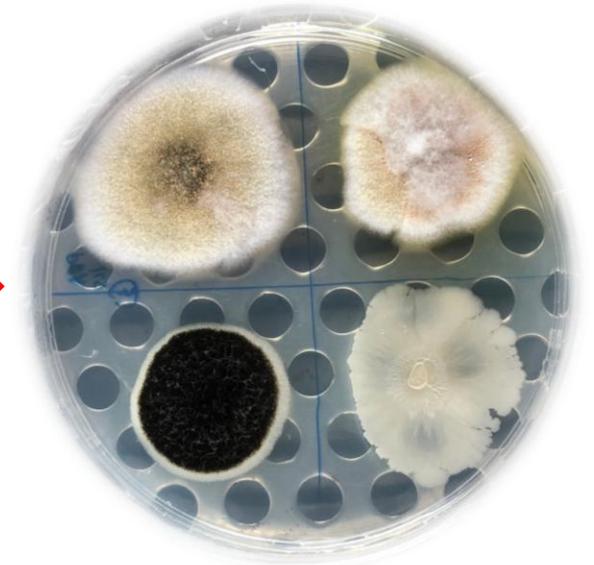
10 Corn husks



20 Others



Cultured in PDA and incubated at 37°C for 5 days



Pure culture in PDA and incubated at 37°C for 5 days

Morphological Identification

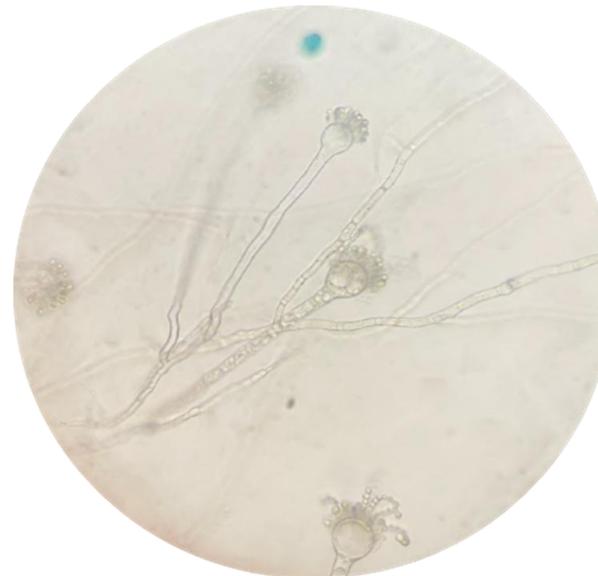
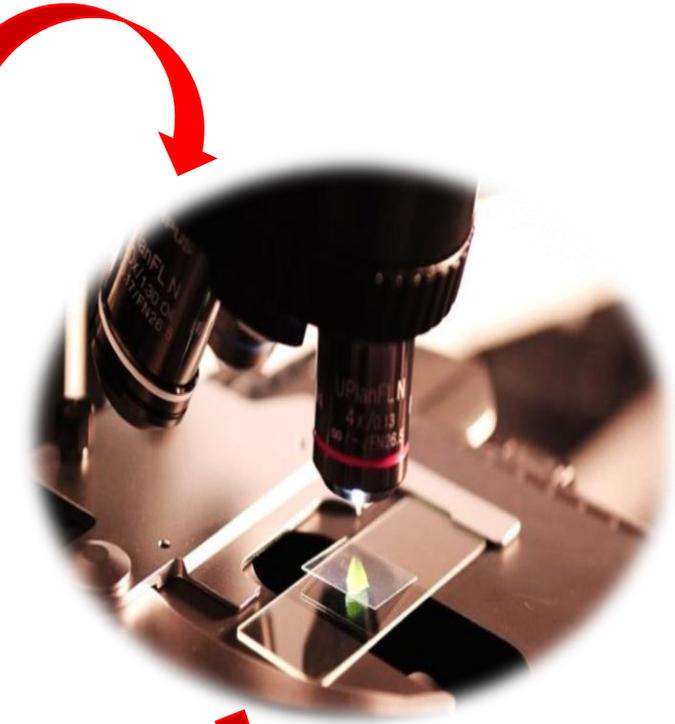
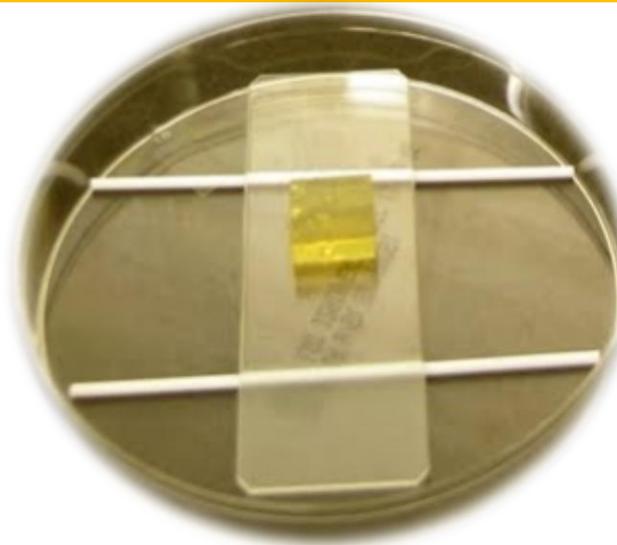
Transferred growing fungi from samples

Cultivated fungi PDA medium

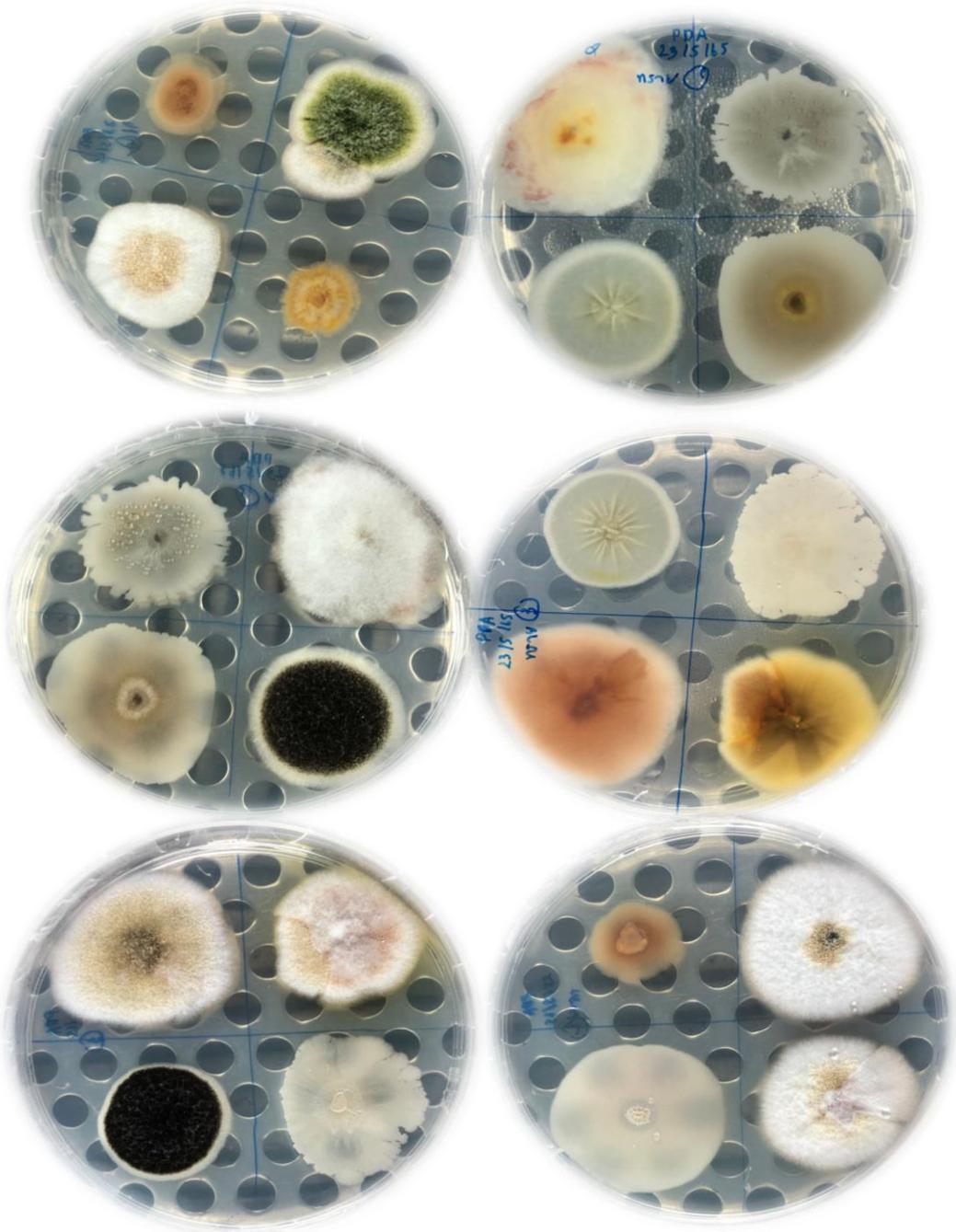
Incubated for 5 days at 37 °C

Checked the fungal culture using
morphological data

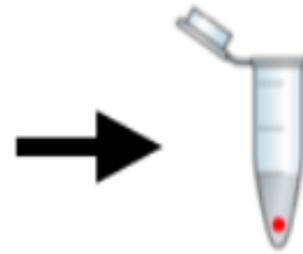
Observed morphology under compound
microscope



DNA extraction and PCR amplification

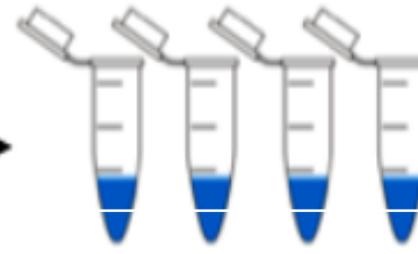


+ 100 μ l Lysis Buffer
+ 2 μ l Enzyme Mix



1. 65°C, 20 min
2. 95°C, 5 min

+ 1 - 2 μ l Lysate



EUDirect 2X PCR Master Mix

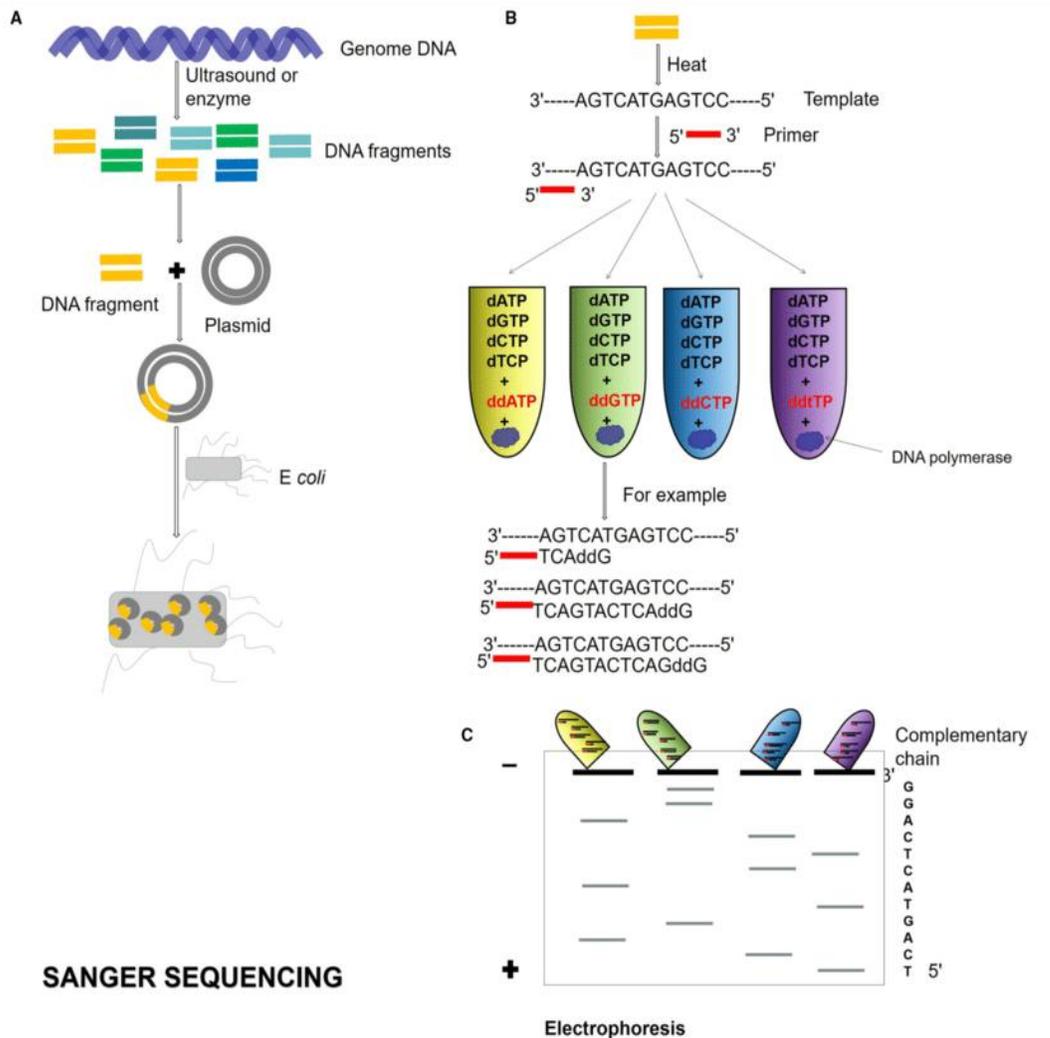


Primers for Detection: **18S rRNA**,
TTS1/TTS4, and **β -tubulin**

The amplification:
95°C.....10 min
45 cycles of
95°C.....15 sec
53°C.....15 sec
72°C.....30 sec

Sequencing

Chain termination method of DNA sequencing



➤ It involves following components:

- Primers (18sRNA, ITS1/ITS4 and β -tubulin)
- DNA template
- DNA polymerase
- dNTPs (A, T, G, C)
- ddNTPs

➤ 4 steps:

- Denaturation
- Primer attachment and extension of bases
- Termination
- Poly acrylamide gel electrophoresis

Phylogenetic analyses

Selection of organisms or a gene family



Choosing appropriate molecular markers



Amplification, sequencing, assembly



Alignment



Evolutionary model



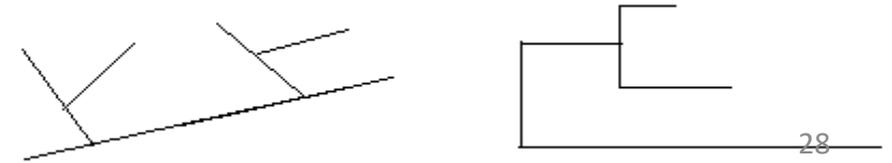
Phylogenetic analysis



Tree construction



Evaluation of phylogenetic tree



Part II: Development and Method validation

LC/MS/MS



Solid-Liquid Extraction Technique
Adapted from the method reported
by Pena *et al.* (2010)

Weigh 2 g of complete feed or corn husks



Add 2 mL distilled water + 8 mL dichloromethane

Vortex for 1 min



Horizontal shaking for 45 min

Sonicate for 10 min



Centrifuge at 4,500 rpm, 15 min, 4°C



Collect supernatant



Filter through Whatman No.4



Evaporate 2.5 mL under nitrogen at 40°C



Reconstitute with 200 µL mobile phase



Filter with 0.22 µm nylon syringe filter, Transfer to LC vial



Analyze by LC-MS/MS



Method validation by LC/MS/MS

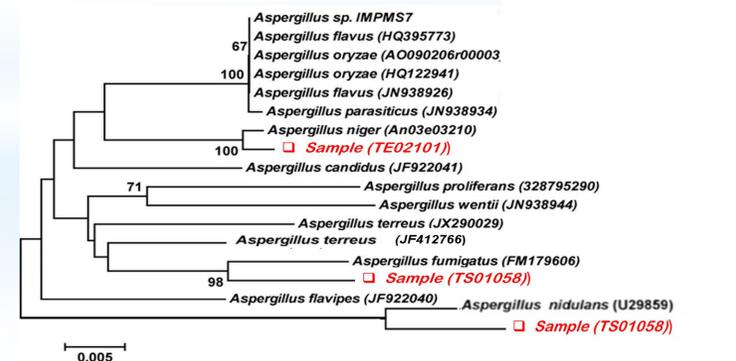
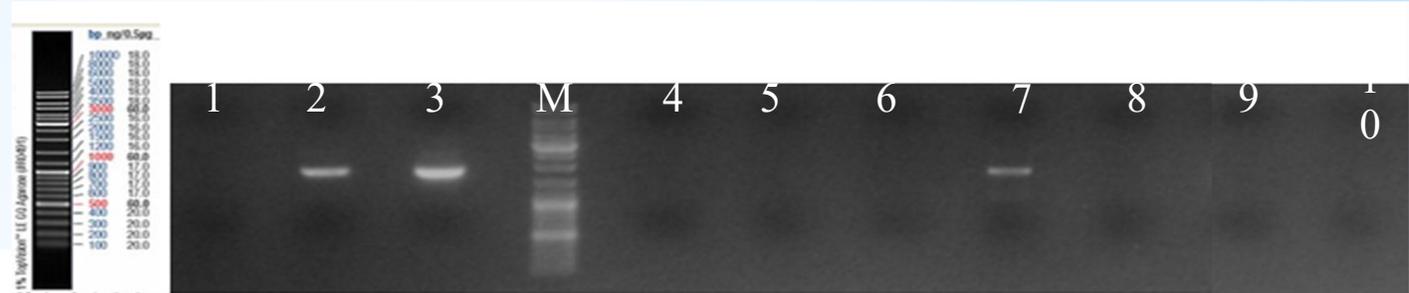
- ❖ **Linearity and range**
- ❖ **Accuracy & precision**
- ❖ **Limits of detection (LOD)**
- ❖ **Limits of quantification (LOQ)**
- ❖ **matrix effect** → **matrix-matched calibration curve**

The validation criteria are based on the guidelines by the European Union, specifically SANTE 2016 and SANTE 2021



Results of Part I

Fungal isolation and molecular characteristics of fungi



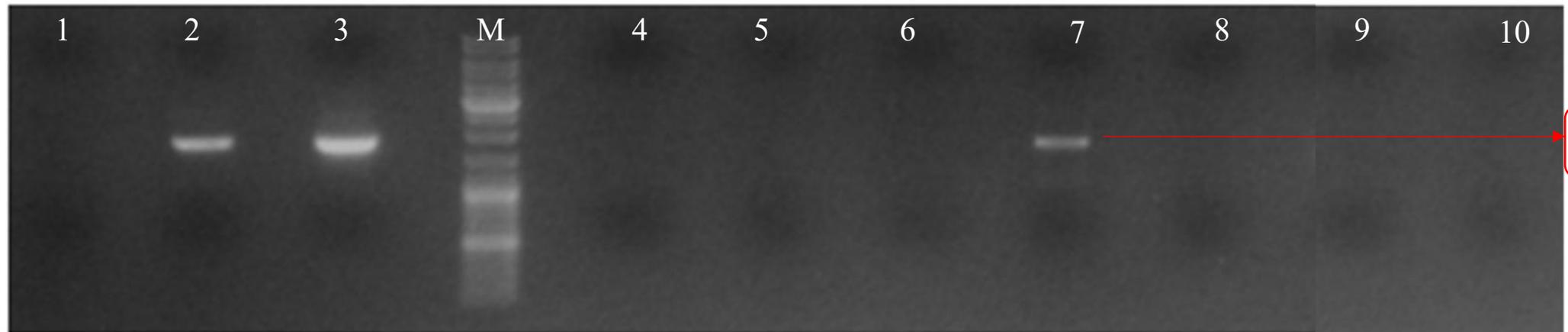
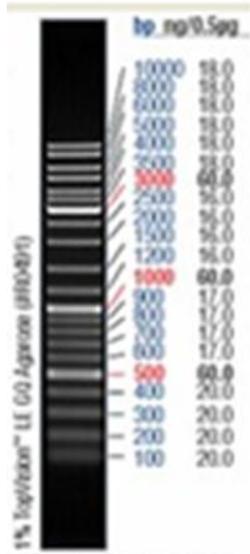
Isolation of different fungi from complete feeds and corn husks (n = 40)

Samples	Fungal species		
	<i>Aspergillus</i> spp.	<i>Monascus</i> spp.	<i>Rhizopus</i> spp.
complete feeds	9/10	5/10	4/10
corn husks	10/10	3/10	5/10



20 Others; Non-fungal species

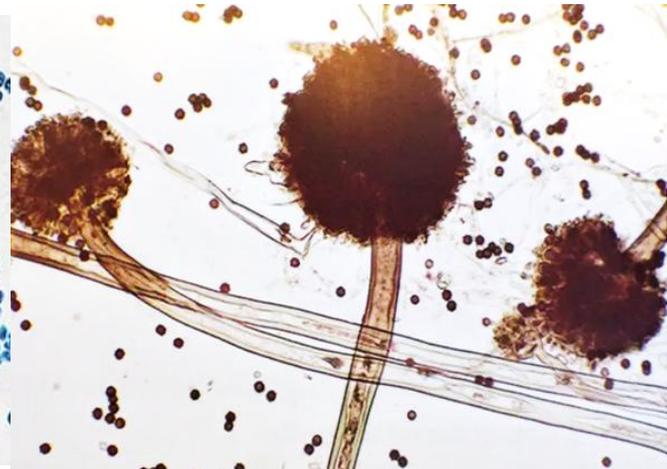
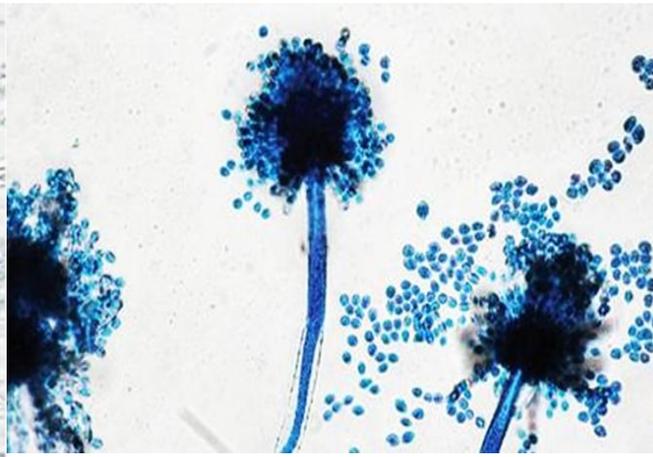
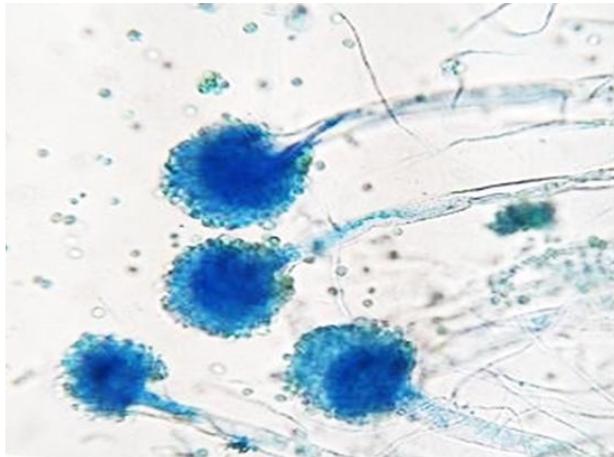
Isolation of *Aspergillus* spp. from complete feeds and corn husks using 18S rRNA primer

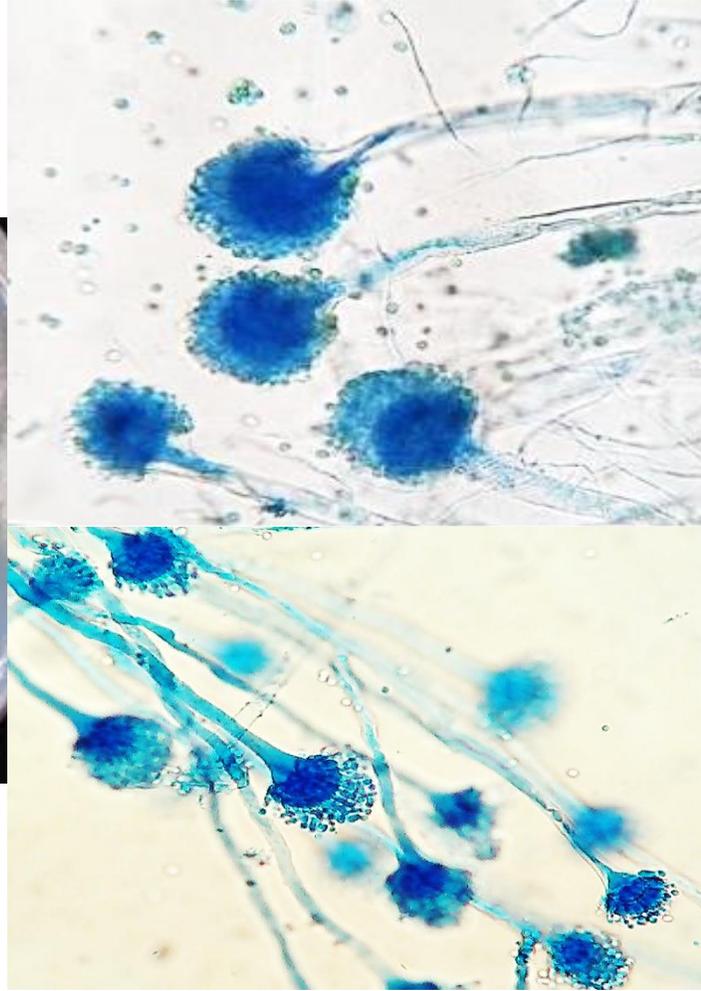
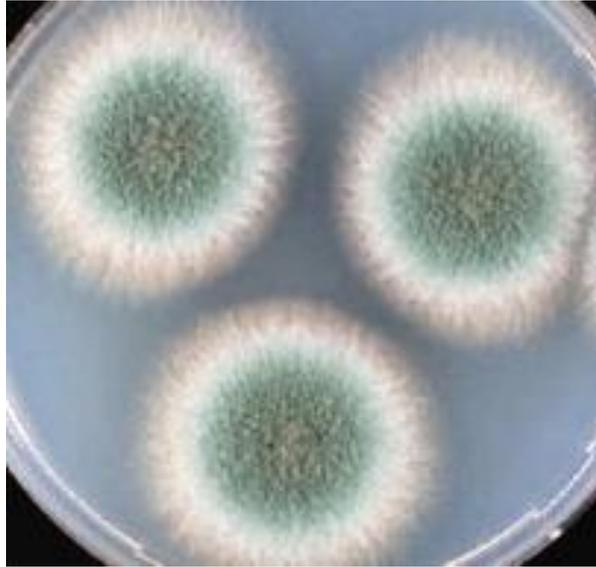


1800 bp

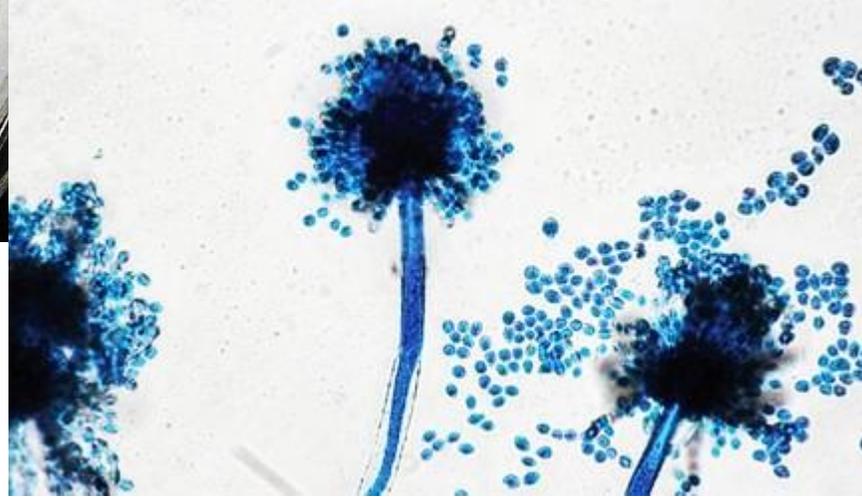
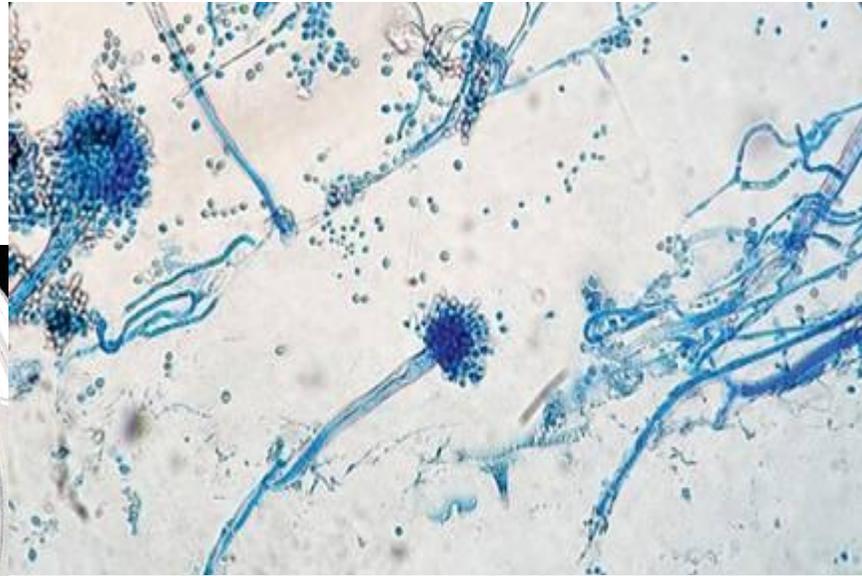
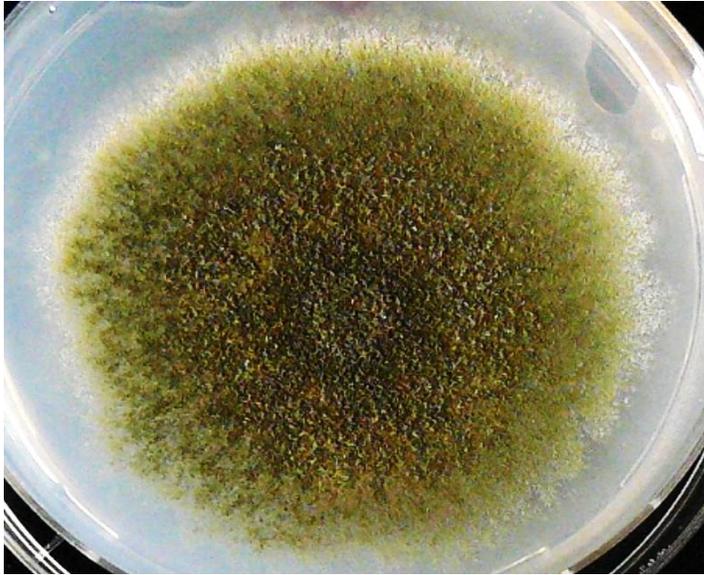
Isolation of *Aspergillus* spp. from complete feeds and corn husks

Samples	Fungal Species		
	<i>Aspergillus fumigatus</i>	<i>Aspergillus nidulans</i>	<i>Aspergillus niger</i>
complete feeds	7/9	3/9	6/9
corn husks	6/10	4/10	3/10

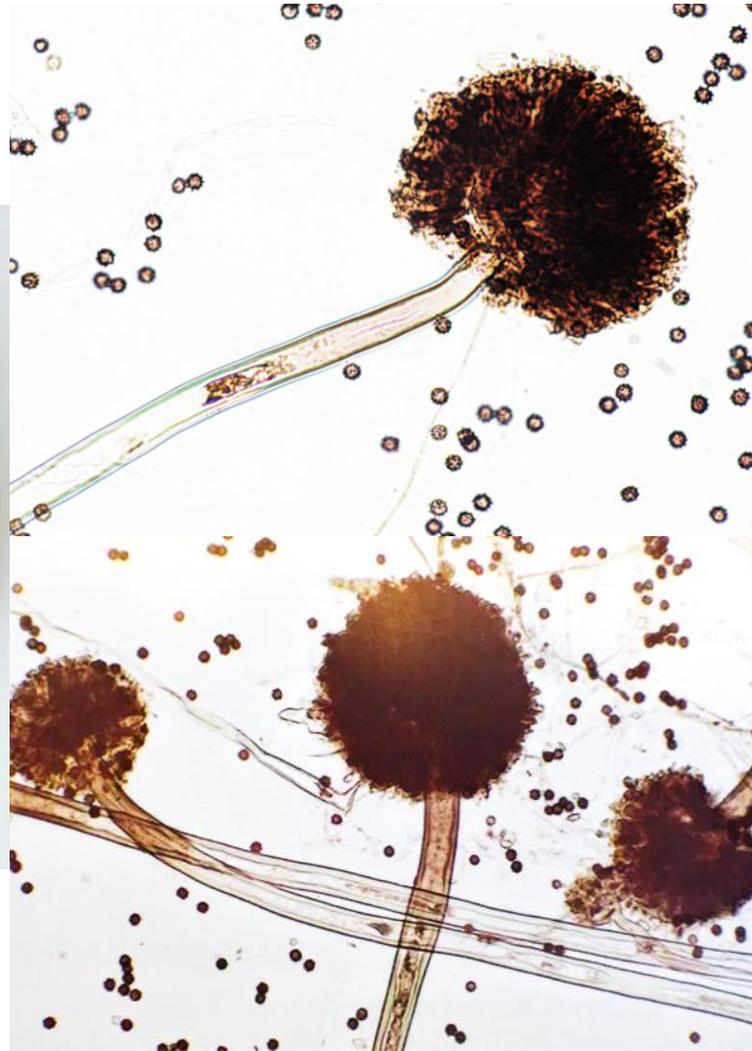




Aspergillus fumigatus

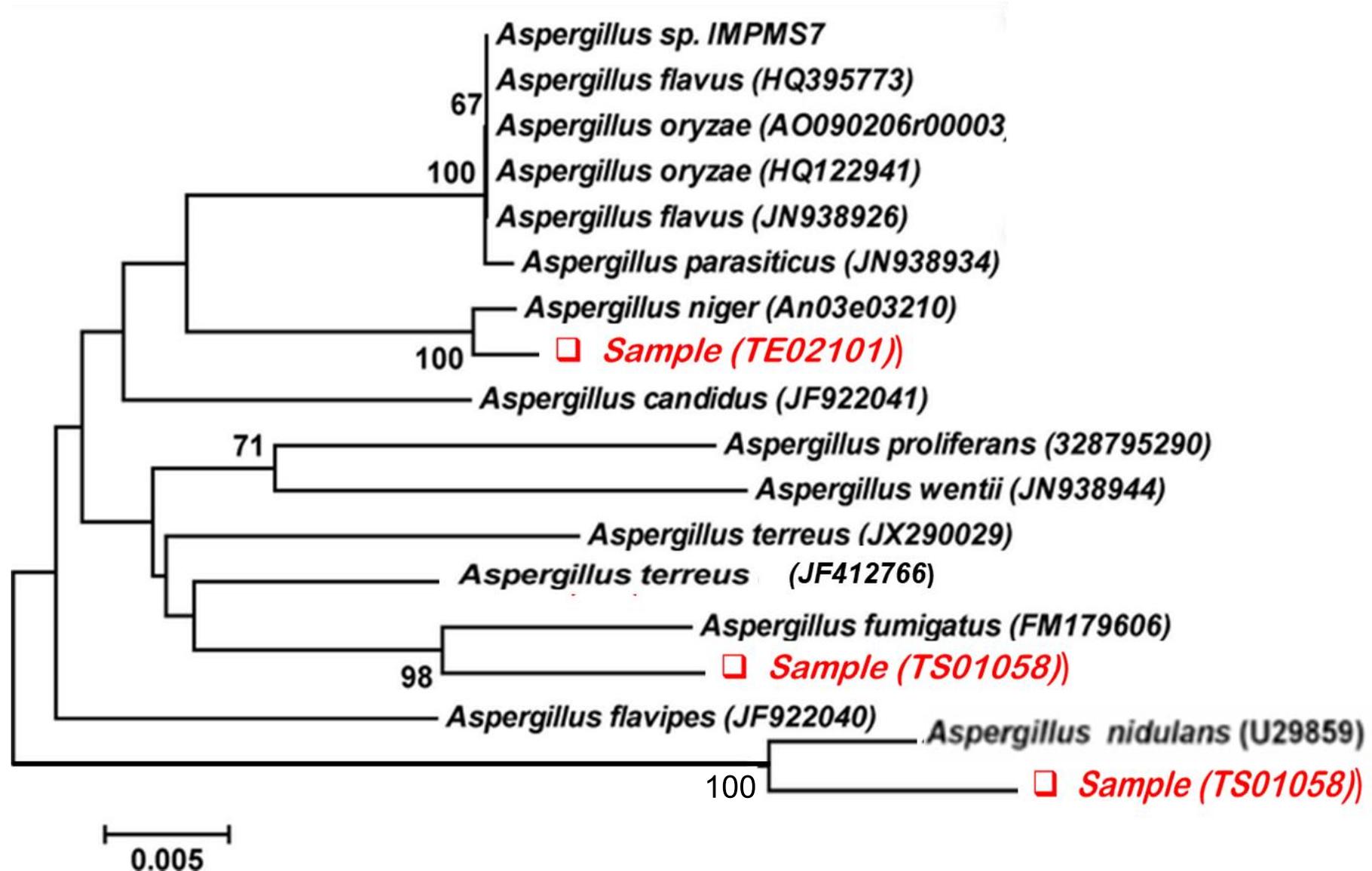


Aspergillus nidulans

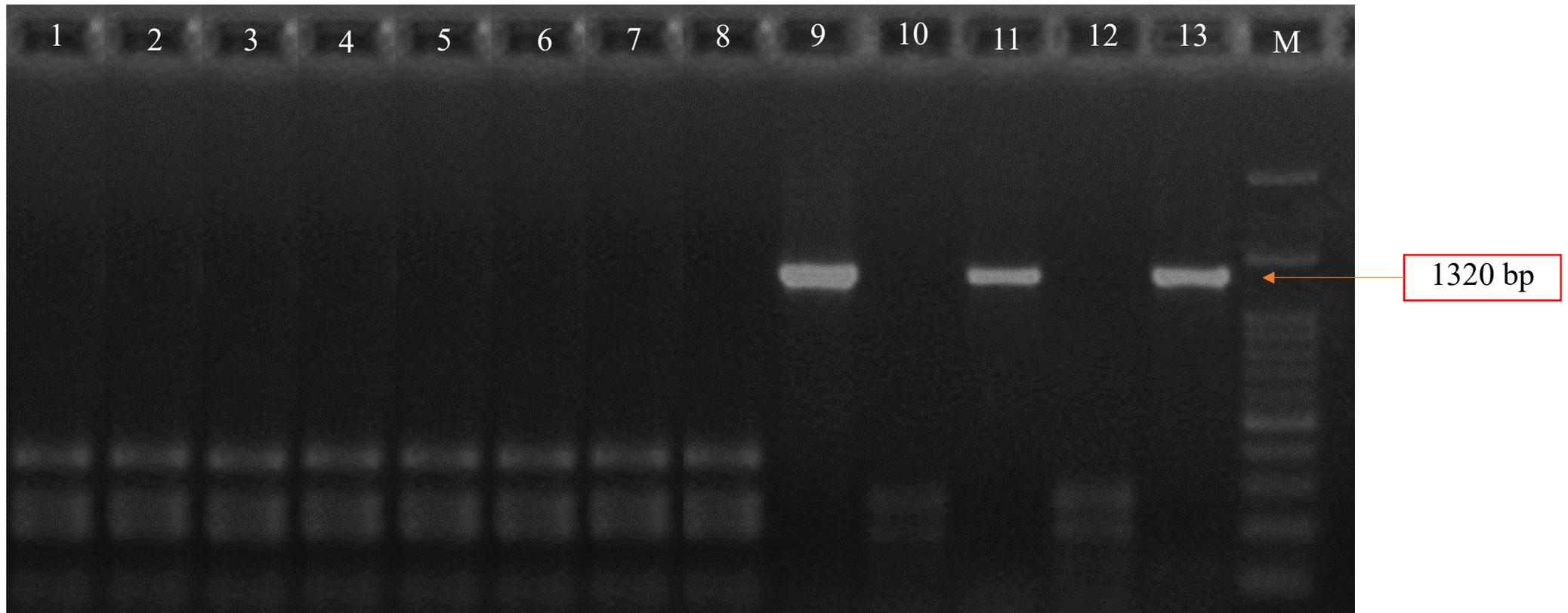


Aspergillus niger

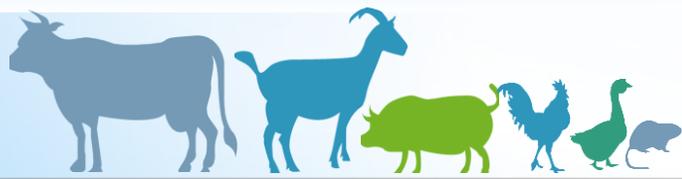
Phylogenetic tree of *Aspergillus* spp.



Agarose gel electrophoresis from *Aspergillus fumigatus* of gliotoxin gene (*gliP*) producing fungi



Results of Part II LC/MS/MS



Mycotoxin analysis

Table 1 HPLC condition for AFB₁ AFB₂ AFG₁ and AFG₂ analysis

Parameters	Information
HPLC Column	Zorbax Eclipse Plus C18 column (50 x 4.6 mm, 1.8 μm)
Column temperature	40° C
Mobile phase	<u>Phase A</u> : 5 mM ammonium formate in 0.2% formic acid in milli-Q <u>Phase B</u> : 0.2% formic acid in ACN
Injection volume	5 μL
Elution mode	Gradient elution
Flow rate	0.4 mL/min
Run time	12 minutes

Table 2 HPLC condition for gliotoxin analysis

Parameters	Information
HPLC Column	Zorbax Eclipse Plus C18 column (50 x 4.6 mm, 1.8 μm)
Column temperature	40° C
Mobile phase	<u>Phase A</u> : 0.1% formic acid in Milli-Q <u>Phase B</u> : 0.1% formic acid in ACN
Injection volume	5 μL
Elution mode	Gradient elution
Flow rate	0.4 mL/min
Run time	12 minutes

Mass analyzer (MS/MS) condition for AFB₁ AFB₂ AFG₁ AFG₂ and gliotoxin analysis

Parameters	Information
Ion source	Electrospray ionization (ESI), positive mode
Gas temp (°C)	325
Gas flow (l/min)	10
Nebulizer (psi)	50
Sheath gas heater	350
Sheath gas flow	11
Capillary (V)	4,000

Mass analyzer (MS/MS) condition for AFB₁ AFB₂ AFG₁ AFG₂ and gliotoxin analysis

Mycotoxins	Precursor ion (m/z)	Product ion (m/z)	Fragmenter (V)	CE (eV)	Polarity	RT (min)
AFB₁	313.1	285.1*	160	21	Positive	7.34
		241	160	35		
AFB₂	315.1	287.1*	160	25	Positive	6.92
		259	160	29		
AFG₁	329.1	311	160	21	Positive	6.90
		243*	160	25		
AFG₂	331.1	313*	160	25	Positive	6.50
		245	160	29		
Gliotoxin	327.1	263.2*	100	0	Positive	6.87
		245.3	100	11		

Remarks : RT = retention time * = quantifier ion CE= Collision energy

The results of the linearity and range of the analytical method and the matrix effect (SSE%) test as determined from tests conducted on complete feed and corn husks.

Mycotoxins	Calibration range (µg/kg)		Correlation coefficient (r)		SSE %	
	Complete feed	Corn husks	Complete feed	Corn husks	Complete feed	Corn husks
AFB ₁	2.0-20.0	2.0-20.0	0.995	0.997	-21.73	-12.03
AFB ₂	2.0-20.0	2.0-20.0	0.998	0.996	-25.18	-14.15
AFG ₁	2.0-20.0	2.0-20.0	0.995	0.999	-18.45	-11.64
AFG ₂	2.0-20.0	2.0-20.0	0.993	0.997	-22.37	-12.21
Gliotoxin	10.0-300	10.0-300	0.996	0.998	-28.65	-15.57

$r \geq 0.990$ (SANTE, 2016; AOAC, 2019)

**A positive %SSE indicates signal enhancement due to the matrix.
A negative %SSE indicates signal suppression caused by the matrix.**

Accuracy and precision of aflatoxins and gliotoxin in complete feed

mycotoxins	Spiked concentration (µg/kg)	accuracy Recovery (%)	Precision (%RSD)	
			Intra-day	Inter-day
AFB ₁	2.0	104.6	5.2	6.4
	10.0	93.7	4.8	7.3
	20.0	90.5	5.1	6.8
AFB ₂	2.0	101.8	7.8	4.6
	10.0	92.7	4.5	8.9
	20.0	91.5	3.1	4.8
AFG ₁	2.0	90.8	5.6	6.1
	10.0	91.5	3.9	4.7
	20.0	96.4	4.0	8.8
AFG ₂	2.0	88.7	6.7	4.6
	10.0	90.2	4.3	5.1
	20.0	89.4	5.1	5.8
Gliotoxin	10	84.5	7.4	8.2
	50	89.9	6.6	7.4
	200	90.6	4.8	6.3

Recovery = 70-120% / %RSD < or = 20% (SANTE, 2021)

Accuracy and precision of aflatoxins and gliotoxin in corn husks

mycotoxins	Spiked concentration (µg/kg)	accuracy Recovery (%)	Precision (%RSD)	
			Intra-day	Inter-day
AFB ₁	2.0	97.5	5.6	8.2
	10.0	95.8	6.1	5.9
	20.0	100.8	4.5	6.3
AFB ₂	2.0	103.3	3.8	5.7
	10.0	105.7	5.5	4.2
	20.0	97.8	2.9	5.5
AFG ₁	2.0	97.5	4.6	6.1
	10.0	101.7	3.0	6.5
	20.0	96.4	4.9	6.7
AFG ₂	2.0	89.8	4.2	4.3
	10.0	90.5	3.3	6.8
	20.0	93.4	6.7	6.4
Gliotoxin	10	90.7	6.4	8.6
	50	92.5	5.6	4.9
	200	90.4	4.4	7.5

Recovery = 70-120% / %RSD < or = 20% (SANTE, 2021)

The limits of detection (LOD) and limits of quantification (LOQ) for the fungal toxins AFB₁, AFB₂, AFG₁, AFG₂, and gliotoxin in complete feed and corn husk samples.

Mycotoxins	complete feed		corn husks	
	LOD (µg/kg)	LOQ (µg/kg)	LOD (µg/kg)	LOQ (µg/kg)
AFB₁	0.65	2.0	0.65	2.0
AFB₂	0.65	2.0	0.65	2.0
AFG₁	0.65	2.0	0.65	2.0
AFG₂	0.65	2.0	0.65	2.0
Gliotoxin	3.2	10.0	3.2	10.0

n = 10, cal. SD

LOD = 3SD

LOQ = 10SD

Concentrations of aflatoxins and gliotoxin in complete feeds (n=9) and corn husks (n=10)

Mycotoxin	complete feeds				corn husks		
	Positive sample (%)	Concentration (µg/kg)		Positive sample (%)	Concentration (µg/kg)		
		Range	Mean		Range	Mean	
AFB₁	66.67	4.11-27.89	15.15	50.0	8.45–24.68	14.07	
AFB ₂	ND	-	-	ND	-	-	
AFG ₁	ND	-	-	ND	-	-	
AFG ₂	ND	-	-	ND	-	-	
Gliotoxin	ND	-	-	ND	-	-	

ND = Not detected (lower than LOD)

Conclusion (*cont.*)

❖ Fungal Contamination in Samples

- ✓ Detected *A. fumigatus*, *A. nidulans*, and *A. niger* in complete feeds and corn husks
- ✓ Only *A. fumigatus* is relevant due to its ability to produce gliotoxin
- ✓ Detection rate of *A. fumigatus* : 77.8% (7/9) in complete feed, 60% (6/10) in corn husks

❖ Aflatoxigenic Fungi

- ✓ No detection of *A. flavus* and *A. parasiticus* from Total samples tested: 19 (complete feed + corn husks)



Conclusion (*cont.*)

Mycotoxin Detection Results

❖ **AFB₁ detected:**

✓ **Complete feed: 4.11–27.89 ppb**

✓ **Corn husks: 8.45–24.68 ppb**

❖ **AFB₂, AFG₁, AFG₂, and GT: Not detected**



Conclusion (*cont.*)

Possible Explanation for Toxin–Fungus Mismatch

- ❖ **AFB₁ found in complete feed and corn husks despite absence of *A. flavus* / *A. parasiticus***
- ❖ **Possible reason:**
 - ✓ **The fungus is dead.**
 - ✓ **AFB₁ remains stable:**
 - ❑ **Resistant to heat >250°C**
 - ❑ **Stable at pH 3–10**



Conclusion (*cont.*)

Absence of Gliotoxin Despite *A. fumigatus*

- ❖ No GT detected in samples
- ❖ Possible environmental limitations:
 - ✓ Temperature
 - ✓ Humidity
 - ✓ Growth phase

Detection of GT Biosynthesis Gene (*gliP*)

- ✓ *A. fumigatus* isolates tested by PCR 13 samples
- ✓ *gliP* gene found in 3 isolates:
 - ❑ 2 from complete feed
 - ❑ 1 from corn husks
- ✓ Presence of *gliP* suggests potential genetic capacity for GT biosynthesis



Conclusion (*cont.*)

- ❖ Isolates positive for the *gliP* gene are genetically capable of GT biosynthesis.
- ❖ If environmental conditions become optimal (suitable temperature, humidity, nutrient availability, and appropriate growth stage), these isolates could potentially produce GT and contaminate feed.



Conclusion (*cont.*)

Sample (n)	<i>Aspergillus flavus, A. parasiticus</i>	AFB ₁ level Concentration (µg/kg)
Complete feed (9)	ND	4.11-27.89
Corn husks (10)	ND	8.45–24.68

ND = Not detected

❖ **AFB₂, AFG₁, & AFG₂: Not detected**

All aflatoxin contaminations were lower than the acceptable level of Animal Feed Control Act. B.E.2018.

Conclusion (*cont.*)

Sample (n)	<i>Aspergillus fumigatus</i>	GT level Concentration (µg/kg)	<i>gliP</i>
Complete feed (9)	7	ND	2
Corn husks (10)	6	ND	1

ND = Not detected (lower than LOD)

Conclusion (*cont.*)

Analytical Method Validation

- ❖ **LC/MS/MS method**
- ❖ **Method validation confirmed: High accuracy and precision**
- ❖ **High sensitivity for low-level detection (ppb range)**
- ❖ **Suitable for quantifying: Aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂), Gliotoxin (GT)**



Conclusion (*cont.*)

Aflatoxins and Gliotoxin Contamination in Dairy Cattle Feed in Bangkok

- ❖ **Low levels of Aflatoxins (AFB1, AFB2, AFG1, AFG2) were found**
- ❖ **All AFs levels were below national safety limits**
- ❖ **Gliotoxin (GT) was not detected in any sample**
- ❖ **Indicates good safety of feed in the Bangkok dairy farming area**
- ❖ **Likely due to effective farm management practices**



Recommendations

- ❖ **Continuous surveillance of AFB1 and GT is essential**
- ❖ **Expand the study to other provinces with significant dairy farming**
- ❖ **Support a larger-scale survey for a more representative data set**



Limitations

- ❖ **The current study is preliminary**
- ❖ **Limited sample size = Not sufficient for national risk assessment**
- ❖ **Cannot be used to set regulatory limits**



Contributions of the Study

- ❖ **Provides a validated LC-MS/MS method**
- ❖ **Can be used as a reliable tool for AFs and GT detection**
- ❖ **Lays a foundation for future monitoring and research**



**Thank You
For Your Attention**

